

Regenerative Biomedicine

Production and Hosting: Shahid Sadoughi University of Medical Sciences



Review Article

Harnessing BMPs for Bone Regeneration: Mechanisms, Biomaterials, and Clinical Applications

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Volume:1 Issue no.3

Editor-in-Chief:

Behrouz Aflatoonian Ph.D.



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Abstract

Bone tissue engineering is a rapidly advancing interdisciplinary domain dedicated to the regeneration of skeletal defects through the combined use of stem cells, bioactive growth factors, and engineered scaffolds. Among the growth factors employed, BMPs, members of the TGF- β superfamily, have emerged as critical mediators of osteogenesis. These proteins drive the differentiation of mesenchymal stem cells and activate key signaling cascades such as Smad, MAPK, and Wnt, which collectively facilitate bone formation. However, translating BMP-based therapies into clinical practice involves including overcoming major challenges, precise dose regulation, spatiotemporal delivery, high production costs, and the risk of adverse effects like ectopic ossification. Recent advances in scaffold engineering, particularly the development of smart biomaterials, offer promising strategies for achieving controlled and targeted BMP delivery. This review comprehensively discusses the molecular mechanisms and biological functions of BMPs, evaluates their integration with scaffold technologies, and examines current clinical applications, limitations, and future directions in the context of bone tissue engineering.

Keywords: Bone tissue engineering, Bone morphogenetic proteins, BMPs, Biomaterial scaffold, Osteogenesis.



How to cite this article: Kuchakzade, F. Harnessing BMPs for Bone Regeneration: Mechanisms, Biomaterials, and Clinical Applications. Regenerative Biomedicine, 2025; 1(3): 235-249.





Introduction

Tissue Engineering is an interdisciplinary field that combines biology, materials science, and engineering to design and construct scaffolds aimed at replacing, repairing, or enhancing the function of damaged or lost tissues in the body. This field has seen significant growth in recent decades, offering various solutions for the treatment of bone, cardiac, liver, neural, and other injuries (1). Among these, bone regeneration stands as one of the most challenging areas of tissue engineering due to the fact that bone is a dynamic, living tissue that, in addition to its structural role, also participates in calcium homeostasis, hematopoiesis, and metabolism (2).

In the design of biological systems for tissue regeneration, the presence of growth factors is essential, in addition to biomaterial scaffolds and stem cells. These factors act as biochemical messengers, guiding cellular differentiation, proliferation, and migration pathways (3). Of the hundreds of identified growth factors, Bone Morphogenetic Proteins (BMPs) are one of the most significant and effective groups for inducing osteogenesis (4). BMPs are a group of secreted proteins that were first discovered by Urist in 1965, who demonstrated that demineralized bone powder could induce new bone formation in non-bony areas (5). It was later determined that this property was due to the presence of BMPs. BMPs are a part of the larger Transforming Growth Factor- β (TGF- β) family and play a critical role in inducing osteogenesis, differentiating mesenchymal stem cells (MSCs), and tissue regeneration (6, 7). BMPs have a unique biological ability to induce osteogenesis (8).

BMP-2 and BMP-7 are two well-known members that have wide applications in bone regeneration and have been approved for use in certain FDA-approved pharmaceutical products (9). These factors can guide stem cells at the injury site towards osteogenesis without the need for external cell supplementation (10). Their functionality is also dependent on the presence of a suitable biomaterial scaffold and а proper microenvironment. Despite the widespread use of BMPs in tissue engineering, challenges such as correct dosing, controlled release, high cost, and potential side effects (such as ectopic bone formation) remain (10, 11). Furthermore, advancements such as nanotechnology, gene therapy, and smart scaffolds could help address these issues (12). Therefore, a comprehensive and systematic review of the various aspects of BMPsranging from biological to technological-can significantly aid in the design of more effective bone regeneration strategies. This review article aims to investigate the following aspects:

1) Molecular Biology and Signaling Pathways of BMPs

2) Role of BMPs in Cellular Differentiation and Induction of Bone Formation

3) Applications in Tissue Engineering Scaffolds

4) Clinical and Preclinical Studies

5) Challenges and Limitations

6)Emerging Technologies for Optimizing BMP Performance

BMPs structure, molecular properties and signaling pathways

BMPs are a subset of the TGF- β family, characterized by a biologically active domain with a conserved structure. BMPs are initially

synthesized as inactive precursors, which include a signal domain, a propeptide, and an active region. After cleavage by proteases, the active fragment is released as a dimer (homo or hetero) in the extracellular space. This dimeric structure is essential for biological activity and is stabilized by disulfide bonds between the monomers (13, 14). The BMP family consists of over 20 different isoforms, which are structurally precursors that transform into active dimeric proteins upon translation (15). BMP-2, BMP-4, and BMP-7 are the most significant members of this family, known for their substantial effects on osteogenesis and differentiation of MSCs (16).

BMPs exert their function by binding to two types of transmembrane serine/threonine kinase receptors: Type I receptors such as Activin receptor-like kinase 2 (ALK2), Activin receptor-like kinase 3 (ALK3), Activin receptor-like kinase 6 (ALK6), and Type II receptors such as BMP receptor Type II (BMPR-II), Activin Receptor Type II (ActR-II), and Activin Receptor Type IIB (ActR-IIB). BMP initially binds to Type II receptors, leading to the activation of the Type I receptor complex, which results in the phosphorylation of downstream signaling proteins (17). Proteins like Endoglin and Betaglycan act as co-receptors for BMPs and play a crucial role in modulating the intensity and type of cellular response. These coreceptors can either enhance or inhibit BMP activity under certain conditions (18).

Smad) signaling pathway is one of the primary pathways involved in BMP action, functioning through the phosphorylation of Smad1, Smad5, and Smad8. This pathway plays a key role in the differentiation of stem cells into osteoblasts, regulation of osteogenic gene expression, and tissue regeneration. Upon BMP binding to the Type I and II receptor complex, the Type I receptor is activated. leading to the direct phosphorylation of Smad1, Smad₅, and Smad8 (12). These phosphorylated Smads then bind to Smad4 (a Co-Smad), forming a complex that translocates to the cell nucleus, where it activates or suppresses the expression of target genes. Key genes activated by this pathway include Runtrelated transcription factor 2 (RUNX2) (a key factor in osteogenesis), Osterix (SP7), and Collagen type I alpha 1 chain (Col1a1), all of which are involved in osteoblast differentiation and extracellular bone matrix formation (19, 20).

tissue **BMP-induced** In engineering, activation of the Smad pathway in MSCs leads to differentiation into osteoblasts. Studies have shown that loading BMPs onto biological scaffolds activates this pathway and enhances osteogenesis in vivo (21). In addition to the classic Smad pathway, BMPs also activate non-Smad signaling can known "non-canonical" pathways, as pathways. These include Mitogen-Activated Protein Kinase (MAPK), Phosphoinositide 3-Kinase/Protein Kinase B (PI3K/Akt), and interactions with the Wingless/ β -catenin (Wnt/ β -catenin) pathway, all of which play significant roles in regulating cell survival, proliferation, migration, and differentiation (22). BMPs can activate the MAPK pathway via Type I receptors. This pathway includes three main branches:

p38 MAPK (increases osteoblast differentiation and expression of genes like Runx2)



Extracellular signal-Regulated Kinases (ERK1/2) (primarily involved in cell proliferation, and in some cases negatively interacts with Smads)

c-Jun N-terminal kinase (JNK) (activated under oxidative stress, with a dual role in cell survival) (23) (Fig. 1).

The PI3K/Akt pathway is especially important for the survival and resistance of stem cells. BMPs activate this pathway via receptor binding and interaction with other factors like Insulin-like growth factor (IGF), which leads to the phosphorylation of Akt. Akt, in turn, enhances cell survival and osteogenic differentiation by inhibiting proteins like Bad and GSK3 β (24). The Wnt pathway, crucial for cell growth and differentiation, interacts with BMP signaling in a complex manner. In many stem cells, simultaneous activation of both the Wnt and BMP pathways enhances osteogenesis. Wnt synergizes with BMP by inhibiting Smad6, which is a negative regulator of BMP Smads (25). In the design of smart scaffolds for bone regeneration, the simultaneous use of factors that activate the MAPK, PI3K, and Wnt pathways alongside BMP can significantly enhance osteoinduction. Additionally, the spatial and temporal control of these pathways is a key to the success of therapeutic outcomes in clinical settings (11, 26). BMP signaling pathways are tightly regulated by a set of negative regulators that prevent overstimulation or inappropriate cellular responses by either inhibiting BMP receptor binding or blocking downstream pathways. These regulators can be divided into two main categories: extracellular inhibitors and intracellular inhibitors (27).

Noggin is one of the most well-known BMP inhibitors, which prevents BMP-2, BMP-4, and BMP-7 from interacting with cell surface receptors by binding directly to them. Noggin plays an inhibitory role in embryonic development, neuronal differentiation, and the regulation of osteogenesis (28). Chordin also inhibits BMP signaling by binding to BMPs and sequestering them away from receptors, through interactions with proteins such as Twisted gastrulation. Chordin is primarily involved in embryonic development and body patterning, but it has also gained importance in tissue engineering (29). Gremlin is an inhibitor involved in both kidney development and the regulation of bone remodeling in adults. This protein is particularly upregulated in pathological conditions like fibrosis and tumorigenesis, exacerbating BMP pathway inhibition (30).

Intracellular Inhibitors

Smad6 and Smad7 (I-Smads), known as inhibitory Smads, prevent the phosphorylation and signal transduction of activated BMP receptors or Smads by binding to them. Notably, Smad6 specifically targets the BMP pathway, inhibiting it in a targeted manner. Some non-canonical pathways, such as ERK, can negatively affect Smads and prevent their translocation to the nucleus, representing a cross-talk mechanism between pathways (27, 31).

In the design of biological scaffolds, the use of BMP inhibitors in specific regions can prevent unwanted ossification in soft tissues. Additionally, controlling the expression of Noggin or Smad6 in biological environments helps achieve more precise and controlled bone regeneration (6).

Extracellular Inhibitors

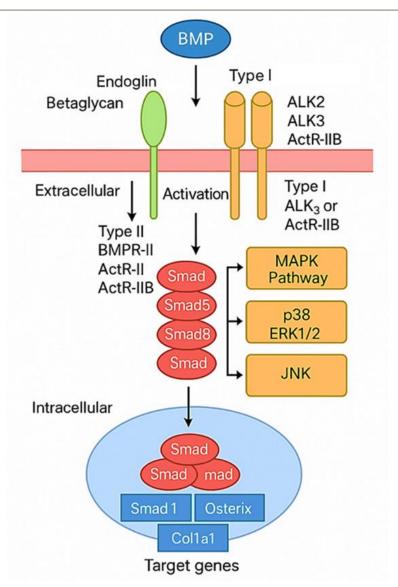


Figure 1. The image depicts a diagram of the BMP (Bone Morphogenetic Protein) pathway and its mechanism of action within the cell. BMP is a morphogenetic growth factor that plays a crucial role in development, cellular differentiation, and cell-cell communication. BMP: Bone Morphogenetic Protein, ALK2: Activin-like Kinase 2, ALK3: Activin-like Kinase 3, ActR-IIB: Activin Receptor Type IIB, BMPR-II: Bone Morphogenetic Protein Receptor Type II, ActR-II: Activin Receptor Type II, Smad: Small Mothers Against Decapentaplegic, MAPK: Mitogen-Activated Protein Kinase, ERK1/2: Extracellular Signal-Regulated Kinase ^{1/}₂, JNK: c-Jun N-terminal Kinase, Col1a1: Collagen Type I Alpha 1.

Application of BMPs in the Design of Bioactive Scaffolds for Bone Tissue Engineering

Bone tissue engineering, as a modern approach to repairing bone defects, is based on three essential pillars: stem cells, growth factors, and biomimetic scaffolds. Among these, scaffolds play a fundamental role as the physical and biochemical substrate for supporting cellular proliferation, differentiation, and organization (32). On the other hand, growth factors, particularly BMPs, have garnered significant attention in



scaffold design due to their unique ability to induce osteogenesis. The aim of advanced scaffold design is to create structures that not only possess suitable physical and mechanical properties but also have the capability for controlled and targeted BMP release (33). Effective scaffolds must be able to deliver and release BMPs efficiently and stably at the site of the bone defect. In Table 1, BMP-carrying biological scaffolds are categorized based on the type of constituent material (10).

Key features of such scaffolds include:

 High biocompatibility: The scaffold should not induce cytotoxicity or immune responses.
 Controlled biodegradability: It should degrade in sync with the formation of new bone, providing space for the regenerating tissue (34).

3) Interconnected porosity: The scaffold must have a porous structure with pore sizes between 100–300 μ m to facilitate cell migration, nutrient exchange, and vascular infiltration (35).

4) Mechanical compatibility: The scaffold should have sufficient strength to withstand mechanical loads and mimic the extracellular matrix (ECM) of bone.

5) Effective BMP loading and release: It must be capable of loading BMPs without deactivation and release them steadily at physiological doses (36).

6) Biological stability: The scaffold should protect BMPs against enzymatic and physiological degradation and release them at the appropriate time (6).

To design BMP delivery strategies in scaffolds, various methods should be considered, including simple physical release, controlled release using biodegradable polymers, BMP-loaded nanoparticles and microspheres, covalent BMP bonding, and multifunctional scaffolds (10, 37).

Simple physical release: In this method, BMPs are adsorbed or loaded onto the surface or bulk of the scaffold. Although it is the simplest approach, it often results in a burst release, leading to a rapid decrease in BMP concentration over time. This method is suitable for short-term applications or when a low dose suffices but is often inadequate for clinical scenarios (38).Controlled many release via biodegradable polymers: Polymers such as Poly (lactic-co-glycolic acid) (PLGA), chitosan, and gelatin can enable sustained BMP release by controlling degradation rates. When combined with ceramics like Beta-Tricalcium Phosphate $(\beta$ -TCP), these scaffolds provide a bone-like environment and protect BMPs from premature degradation. This method has shown excellent results in animal models by stimulating endogenous bone formation (39). Use of BMP-loaded nanoparticles and microspheres: Polymeric or inorganic nanoparticles like PLGA and calcium phosphate (CaP) can encapsulate BMPs and release them at controlled rates depending on physiological conditions. This approach offers enhanced stability, protection against enzymatic degradation, and more precise dosage control (40).

Covalent bonding of BMP to scaffold matrix: Here, BMPs are covalently bound to the scaffold and are only released in the presence of specific enzymes in the target tissue. This strategy enhances targeting, reduces systemic effects, and improves therapeutic efficacy. It is feasible through advanced chemical reactions and responsive systems (41). Multifunctional scaffolds: These scaffolds **Table 1.** Examples of classification of biomimetic scaffolds containing bone morphogenetic protein (BMP) (Abbreviation: BMPs: Bone Morphogenetic Proteins, PLGA: Poly (lactic-co-glycolic acid), PCL: Polycaprolactone, HA: Hydroxyapatite, β-TCP: Beta-Tricalcium Phosphate, GelMA: Gelatin Methacrylate)

	Scaffold Type	Examples	Advantages	Disadvantages
1	Natural	Collagen, Chitosan	Biocompatible, bioactive	Rapid degradation, low strength (42)
2	Synthetic	PLGA, PCL	Controlled degradation and structure	Requires surface modification (43, 44)
3	Inorganic	ΗΑ, β-ΤСΡ	Bioactive, bone-like	Brittle, high release rate (45)
4	Composite	PLGA/HA, GelMA/β-TCP, gelatin/HA/β-TCP, PCL/PLGA/β-TCP	Combines benefits	Complex manufacturing (46-48)

offer additional functionalities alongside BMP delivery, such as electrical conductivity, antibacterial properties, or co-delivery of other factors like Vascular Endothelial Growth Factor (VEGF) (49). Examples include:

1)PLGA(BMP-2) / CS (Pac-525) @ MC/PCL scaffold: Provides osteogenic and antibacterial effects (50).

2) 3D-printed PCL/bioglass (BGS-7) composite scaffolds: Offers customizable design and high bone regeneration potential (51).

3) Chitosan-loaded mesoporous silica nanoparticle scaffolds containing BMP-2 and dexamethasone: Significant Stimulation of Osteoblast Differentiation and Bone Regeneration by Simultaneous Delivery of BMP-2 and Dex (52).

Bone morphogenetic proteins, especially BMP-2 and BMP-7, are among the few growth factors that have received FDA approval for clinical use in bone regeneration (53). One of the most well-known commercial products in this field is Infuse® Bone Graft, developed by Medtronic, which combines BMP-2 with an absorbable collagen scaffold (54). This product is approved for:

1) Spinal fusion surgeries: Assists vertebral fusion, especially in high-risk patients.

2) Craniofacial bone defect reconstruc-tion reconstruction: Such as in large fractures or post-tumor resection repairs.

3) Orthopedic surgeries: For treating long bone diaphyseal defects, nonunions, and complex fractures (55, 56).

Clinical studies have shown that BMP-2 combined with collagen scaffolds enhances the rate and quality of bone formation, reduces the need for autografts, and facilitates healing in challenging cases (6). Similarly, BMP-2 incorporated into β -TCP scaffolds has been successfully used in alveolar bone reconstruction, yielding improved healing outcomes (57). Moreover, animal and clinical studies have reported that BMP-7-loaded scaffolds are effective in



treating nonunion fractures (58). However, concerns such as ectopic bone formation, excessive inflammation, and potential carcinogenic risks at high doses have been reported (59). Key challenges in BMP-based scaffold applications include:

1) Precise dosage control: High doses may cause ectopic bone or inflammation; low doses may be ineffective (60).

2) BMP stability in moist and enzymatic body environments (61).

3) Variable immune responses among patients (62).

4) Lack of standardization in clinical testing(63).

To address these challenges, emerging technologies such as smart scaffolds sensitive to pH and enzymes for the controlled release of BMP, the use of 3D bioprinting for precise design and spatial loading of BMP, the integration of BMP with exosomes (64-67) to enhance stability and efficacy, and "nanohybrid" scaffolds for synergistic combination of BMP with other factors such as VEGF are currently under investigation (26, 68, 69)

Clinical Studies and Therapeutic Applications of BMPs in Bone Regeneration

BMPs, particularly rhBMP-2 and rhBMP-7, have played a key role in clinical treatments related to bone regeneration in recent years, including complex fractures, nonunions, large bone defects, and spinal and maxillofacial surgeries (10). These biological factors promote the differentiation of mesenchymal stem cells into osteoblasts and enhance osteogenesis, offering an effective alternative to autologous bone grafting. FDAapproved BMPs include the following: 1) rhBMP-2: This factor is used in lumbar interbody fusion, repair of mandibular bone defects, sinus lift in dental surgery, and in combination with absorbable collagen sponge (ACS).

2) rhBMP-7 (OP-1): This factor is used in the treatment of nonunion in long bones and complex orthopedic surgeries, often in combination with biological scaffolds or natural carriers (54, 70).

Currently, these factors are used in spinal surgery, delayed fracture healing, maxillofacial reconstruction, and dental implants (71). In lumbar spinal fusion surgeries, BMPs have served as an alternative to autologous bone grafts and have shown favorable outcomes (72).

In a systematic review and meta-analysis, the efficacy and safety of rhBMP-2 and autologous iliac crest bone graft (ICBG) in lumbar fusion were compared in 2,185 patients. The results showed that rhBMP-2 had a higher fusion success rate, a lower risk of reoperation, and a similar complication rate compared to ICBG. rhBMP-2 was recommended as an effective alternative to ICBG for lumbar fusion (73). In a study, existing research on the application of rhBMP-2 in spinal fusion surgery from 1965 to 2022 was reviewed. The evidence shows that the fusion rate with rhBMP-2 is similar to or even higher than the results achieved with autologous bone grafts. However, there are concerns regarding the cost, optimal dosage, and potential complications associated with the use of rhBMP-2 in spinal surgery (72). In a review study, the effect of rhBMP-2 in maxillofacial surgeries was investigated. The study showed that rhBMP-2 has high potential maxillofacial surgeries due to its in osteoinductive properties. Studies have indicated that rhBMP-2 can reduce donor site

morbidity and increase bone height in sinus and ridge augmentation procedures. Additionally, in the treatment of medicationrelated osteonecrosis of the jaw, rhBMP-2 has been used as an adjunct with promising results. Overall, rhBMP-2 is recognized as a promising graft material in maxillofacial surgery (74).

In a study, the safety and efficacy of local rhBMP-7 implantation in bovine-derived collagen paste for the treatment of resistant non-unions were evaluated in fifty-two patients. The results showed that 94% of patients achieved union, with a mean healing time of 5.6 months. This method was recognized as an effective adjunctive treatment, with only one case of synostosis reported as a complication (75). A study evaluated the treatment of nonunion of the femur using single-stage methods (with rhBMP-7 and RIA) and two-stage methods (Masquelet technique). A total of 88 patients were assessed, with 74% achieving bone consolidation on average within 9.3 months. Intramedullary reaming (83%) was more effective than osteosynthesis plates (60%), and smoking was associated with a reduced of consolidation. rate The Masquelet technique was effective for infections and large bone defects, and quality of life improved in both groups. Treatment with rhBMP-7 and RIA was recommended for small defects, while intramedullary nailing was advised for large defects (76).

Despite the widespread application of BMPs in clinical settings, side effects such as ectopic bone formation, severe swelling, and immune responses are notable (10). To overcome these side effects and limitations, new research strategies are being explored, such as the use of nanoparticles or hydrogels for controlled BMP release, combining BMP with other factors like VEGF to enhance both osteogenesis and angiogenesis, and gene editing (Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR)) to improve endogenous BMP production by stem cells (77, 78).

Challenges, Innovations, and Future Directions in the Application of BMPs in Bone Tissue Engineering

While BMPs have been proposed as effective biological agents for bone regeneration, various challenges remain in optimizing their use (79). These challenges, ranging from safety concerns and side effects to economic barriers and the need for improved BMP production and release processes, can significantly impact the clinical application of these proteins (80). One common issue with the use of BMPs, especially at high doses, is ectopic bone formation (81). This problem arises from the non-targeted distribution of BMPs to surrounding areas during implantation (82). Bone formation in inappropriate locations such as soft tissues, nerves, or within the spinal cavity can lead to severe complications, including nerve compression and eventual nerve damage (83). The use of smart carriers that can precisely direct BMPs to the intended site and improvements in the design of biodegradable scaffolds to limit BMP release and prevent its spread to undesired areas can be effective solutions to address this issue (41).

In many cases, high doses of rhBMPs are required to achieve effective bone formation. These high doses can lead to severe inflammation, swelling, and immune system reactions (6). In some patients, the immune



response to BMPs may lead to the production of anti-BMP antibodies, reducing the therapeutic effect (84). The use of controlledrelease systems that allow for more precise and gradual BMP delivery, along with reduced BMP doses through optimized scaffolds and carriers, could offer viable solutions to this problem (7). Another major challenge is the precise control of BMP release. Many current carriers, such as collagen sponges, are not capable of targeted and controlled BMP release (36). This issue results in diminished BMP effectiveness over time and increased side effects. Using nanoparticles or hydrogels that offer better control over drug release is a promising solution (85). Additionally, designing smart release systems that can respond to specific environmental conditions, such as pH or temperature, can lead to more effective BMP release (86, 87).

The use of biodegradable nanoparticles, such as PLGA (poly-lactic-co-glycolic acid), and liposomes for the gradual release of BMPs at the target site offers a more accurate and effective means of delivering BMPs to the target tissue, preventing unintended release (6, 88). This approach is associated with reduced side effects and prolonged BMP release at specific locations (89). An exciting innovation is the use of engineered stem cells for local BMP production. These cells can produce BMPs within the patient's body and aid in bone regeneration (90). Gene editing techniques using CRISPR-Cas9 could also be employed to engineer stem cells to enhance BMP production. This in-situ production technology reduces the need for external release and increases therapeutic efficiency (91, 92).

Nano-composite and micro-layered scaffolds that can stimulate cell differentiation into

osteoblasts and precisely control BMP release are also an exciting advancement (1, 93, 94). These scaffolds, using biodegradable materials, allow the body to naturally build new bone structure (95). This technology controls BMP release and creates a favorable environment for bone growth (96). One of the most important areas of progress is combining BMPs with other growth factors such as VEGF or FGF (fibroblast growth factor). These combinations can simultaneously enhance processes like osteogenesis and angiogenesis (Table 2) (97). In recent decades, BMPs have become one of the most widely used tools in bone tissue engineering. These proteins are capable of stimulating bone production and repairing damaged tissues. Recent advances in production, scaffold design, and combination therapies have rapidly expanded the clinical and experimental applications of BMPs (98).

Discussion

BMPs have emerged as pivotal agents in the field of tissue engineering due to their potent osteoinductive capabilities. Their role transcends simple bone induction; they orchestrate complex signaling cascades that mediate stem cell differentiation, matrix synthesis, and angiogenesis, all of which are crucial for effective bone regeneration (99). The canonical Smad pathway remains the most studied route of BMP signaling, especially through BMP-2 and BMP-7, which have received FDA approval for clinical applications (100). These pathways not only induce osteoblast differentiation but also work in concert with non-canonical pathways such as MAPK and PI3K/Akt, creating a robust regulatory network (101). However, BMPs are

Table 2. Combination of BMPs with other growth factors to enhance bone formation and angiogenesis (BMP: Bone Morphogenetic Protein; VEGF: Vascular Endothelial Growth Factor; FGF-2: Fibroblast Growth Factor-2; PDGF: Platelet-Derived Growth Factor. Numbers in parentheses refer to references.)

	Combination	Benefit
1	BMP + VEGF	Simultaneous enhancement of osteogenesis and angiogenesis (102)
2	BMP + FGF-2	Stimulation of osteoblast differentiation and stem cell proliferation (103)
3	BMP + PDGF	Simultaneous repair of soft and hard tissues (104)

not without limitations. High doses are often required to achieve therapeutic efficacy, which can lead to adverse effects such as ectopic bone formation and inflammation (105). This has led to a growing interest in delivery strategies that can precisely control the spatial and temporal release of BMPs. Incorporating BMPs into smart biomimetic scaffolds, such as hydrogels or nanofiber composites, offers a promising solution by mimicking the native extracellular matrix and enhancing bioavailability (106, 107).

Another critical challenge lies in the finetuning of BMP signaling. Inhibitory molecules like Noggin, Gremlin, and Smad6 are essential for preventing aberrant ossification, but they can also hinder therapeutic outcomes if not carefully modulated (108, 109). Emerging technologies, including gene editing and controlled gene expression systems, provide new avenues for overcoming these challenges. combinatorial Furthermore, approaches involving BMPs and other growth factors such as VEGF or IGF may enhance vascularization and support complex tissue repair (109, 110). From a clinical standpoint, although BMPbased therapies have achieved notable success in spinal fusion and long bone defects, translation to broader applications requires addressing concerns over cost, reproducibility

and safety (111). Continuous innovation in scaffold design, molecular biology, and regenerative medicine is therefore essential to maximize the therapeutic potential of BMPs in tissue engineering.

Conclusion

In summary, BMPs play an indispensable role in modern tissue engineering, particularly in bone regeneration. Their unique biological properties, ability to modulate multiple signaling pathways, and synergy with biomaterials position them at the forefront of regenerative strategies. However, to fully harness their potential, several obstaclesincluding dose optimization, delivery precision, and inhibition management-must be addressed. Future research should prioritize interdisciplinary approaches combining molecular biology, materials science, and clinical translational studies. Innovations in smart biomaterials, gene regulation, and combinatorial growth factor therapies hold the key to overcoming current limitations and broadening the clinical application of BMP-based tissue engineering. By bridging fundamental biological mechanisms with advanced scaffold technologies, the next generation of BMPcentered therapies can offer safe, effective,



and personalized solutions for complex tissue

regeneration challenges.

References

1. Farjaminejad S, Farjaminejad R, Garcia-Godoy F. Nanoparticles in bone regeneration: a narrative review of current advances and future directions in tissue engineering. Journal of Functional Biomaterials. 2024;15(9):241.

2. Todd EA, Mirsky NA, Silva BLG, Shinde AR, Arakelians AR, Nayak VV, et al. Functional Scaffolds for Bone Tissue Regeneration: A Comprehensive Review of Materials, Methods, and Future Directions. Journal of Functional Biomaterials. 2024;15(10):280.

3. Setiawan AM, Kamarudin TA, Abd Ghafar N. The role of BMP4 in adipose-derived stem cell differentiation: A minireview. Frontiers in Cell and Developmental Biology. 2022;10:1045103.

4. Rodríguez-Merchán EC. A review of recent developments in the molecular mechanisms of bone healing. International Journal of Molecular Sciences. 2021;22(2):767.

5. Urist MR. Bone: formation by autoinduction. Science. 1965;150(3698):893–9.

6. Qi J, Wu H, Liu G. Novel strategies for spatiotemporal and controlled BMP-2 delivery in bone tissue engineering. Cell Transplantation. 2024;33:09636897241276733.

7. Oliveira ÉR, Nie L, Podstawczyk D, Allahbakhsh A, Ratnayake J, Brasil DL, et al. Advances in growth factor delivery for bone tissue engineering. International journal of molecular sciences. 2021;22(2):903.

8. Guillot-Ferriols M, Lanceros-Méndez S, Ribelles JG, Ferrer GG. Electrical stimulation: Effective cue to direct osteogenic differentiation of mesenchymal stem cells? Biomaterials Advances. 2022;138:212918.

9. Sen A, Qamar R, Choubisa R, Parikh M, Shah D. BMP modulation of osteogenesis: molecular interactions and clinical applications. Proceedings of the Indian National Science Academy. 2025:1–7.

 Zhu L, Liu Y, Wang A, Zhu Z, Li Y, Zhu C, et al. Application of BMP in bone tissue engineering. Frontiers in Bioengineering and Biotechnology. 2022;10:810880.
 Song P, Zhou D, Wang F, Li G, Bai L, Su J. Programmable biomaterials for bone regeneration. Materials Today Bio. 2024:101296.

12. Zou M-L, Chen Z-H, Teng Y-Y, Liu S-Y, Jia Y, Zhang K-W, et al. The Smad dependent TGF- β and BMP signaling pathway in bone remodeling and therapies. Frontiers in molecular biosciences. 2021;8:593310.

13. Akiyama T, Raftery LA, Wharton KA. Bone morphogenetic protein signaling: the pathway and its regulation. Genetics. 2024;226(2):iyad200.

14. Ehata S, Miyazono K. Bone morphogenetic protein signaling in cancer; some topics in the recent 10 years. Frontiers in cell and developmental biology. 2022;10:883523.

15. Halloran D, Durbano HW, Nohe A. Bone morphogenetic protein-2 in development and bone homeostasis. Journal of developmental biology. 2020;8(3):19.

16. Wang Z, Ren L, Li Z, Qiu Q, Wang H, Huang X, et al. Impact of Different Cell Types on the Osteogenic Differentiation Process of Mesenchymal Stem Cells. Stem Cells International. 2025;2025(1):5551222.

17. Xie Z, Zhou G, Zhang M, Han J, Wang Y, Li X, et al. Recent developments on BMPs and their antagonists in inflammatory bowel diseases. Cell Death Discovery. 2023;9(1):210.

18. Pawlak JB, Blobe GC. TGF- β superfamily correceptors in cancer. Developmental Dynamics. 2022;251(1):117–43.

19. Komori T. Regulation of proliferation, differentiation and functions of osteoblasts by Runx2. International journal of molecular sciences. 2019;20(7):1694.

20. Firdauzy MAB, Ahmad NB, Setiawatie EM, Rahmatari B, Roestamadji RI. BMP2 and Osterix Interaction in Osteoblastogenesis: An Article Review. Malaysian Journal of Medicine & Health Sciences. 2024;20.

21. Garcia J, Delany AM. MicroRNAs regulating TGF β and BMP signaling in the osteoblast lineage. Bone. 2021;143:115791.

22. Loh H-Y, Norman BP, Lai K-S, Cheng W-H, Nik Abd. Rahman NMA, Mohamed Alitheen NB, et al. Post-transcriptional regulatory crosstalk between MicroRNAs and canonical TGF- β /BMP signalling cascades on osteoblast lineage: a comprehensive review. International Journal of Molecular Sciences. 2023;24(7):6423.

23. Liu X, Zhao W, Peng Y, Liu N, Liu Q. The relationship between MAPK signaling pathways and osteogenic differentiation of periodontal ligament stem cells: a literature review. PeerJ. 2025;13:e19193.

24. Wang L, Ruan M, Bu Q, Zhao C. Signaling Pathways Driving MSC Osteogenesis: Mechanisms, Regulation, and Translational Applications. International Journal of Molecular Sciences. 2025;26(3):1311.

25. Arya PN, Saranya I, Selvamurugan N. Crosstalk between Wnt and bone morphogenetic protein signaling during osteogenic differentiation. World Journal of Stem Cells. 2024;16(2):102.

26. Jiang S, Wang M, He J. A review of biomimetic scaffolds for bone regeneration: toward a cell-free strategy. Bioengineering & Translational Medicine. 2021;6(2):e10206.

27. Kulikauskas MR, Bautch VL. The versatility and paradox of BMP signaling in endothelial cell behaviors and blood vessel function. Cellular and Molecular Life Sciences. 2022;79(2):77.

28. Ma Y, Bi W. Research Progress of Osteogenesis Induced Jointly by BMP and BGN. International Core Journal of Engineering. 2022;8(1):441–6.

29. Yan Y, Wang Q. BMP signaling: lighting up the way for embryonic dorsoventral patterning. Frontiers in Cell and Developmental Biology. 2021;9:799772.

30. O'Reilly S. Gremlin: a complex molecule regulating wound healing and fibrosis. Cellular and Molecular Life Sciences. 2021;78(24):7917–23.

31. Wu M, Wu S, Chen W, Li Y-P. The roles and regulatory mechanisms of TGF- β and BMP signaling in

bone and cartilage development, homeostasis and disease. Cell research. 2024;34(2):101–23.

32. Ferraz MP. An overview on the big players in bone tissue engineering: biomaterials, scaffolds and cells. International Journal of Molecular Sciences. 2024;25(7):3836.

33. Vermeulen S, Birgani ZT, Habibovic P. Biomaterial-induced pathway modulation for bone regeneration. Biomaterials. 2022;283:121431.

34. Negi A, Goswami K, Diwan H, Agrawal G, Murab S. Designing osteogenic interfaces on 3D-Printed thermoplastic bone scaffolds. Materials Today Chemistry. 2025;45:102635.

35. Mohammadi H, Sepantafar M, Muhamad N, Bakar Sulong A. How does scaffold porosity conduct bone tissue regeneration? Advanced Engineering Materials. 2021;23(10):2100463.

36. Szwed-Georgiou A, Płociński P, Kupikowska-Stobba B, Urbaniak MM, Rusek-Wala P, Szustakiewicz K, et al. Bioactive materials for bone regeneration: biomolecules and delivery systems. ACS biomaterials science & engineering. 2023;9(9):5222–54.

37. Suamte L, Tirkey A, Barman J, Babu PJ. Various manufacturing methods and ideal properties of scaffolds for tissue engineering applications. Smart Materials in Manufacturing. 2023;1:100011.

38. Tilkin RG, Mahy JG, Grandfils C, Lambert SD. Tissue engineering using scaffolds for bone reconstruction: a review of sol-gel silica materials for bone morphogenetic proteins (BMP) encapsulation and release. Journal of Sol-Gel Science and Technology. 2022;103(2):616–36.

39. Batool M, Abid MA, Javed T, Haider MN. Applications of biodegradable polymers and ceramics for bone regeneration: a mini-review. International Journal of Polymeric Materials and Polymeric Biomaterials. 2025;74(1):39–53.

40. Bello SA, Cruz-Lebrón J, Rodríguez-Rivera OA, Nicolau E. Bioactive scaffolds as a promising alternative for enhancing critical-size bone defect regeneration in the craniomaxillofacial region. ACS Applied Bio Materials. 2023;6(11):4465–503.

41. Percival KM, Paul V, Husseini GA. Recent advancements in bone tissue engineering: integrating smart scaffold technologies and bio-responsive systems for enhanced regeneration. International Journal of Molecular Sciences. 2024;25(11):6012.

42. Donnaloja F, Jacchetti E, Soncini M, Raimondi MT. Natural and synthetic polymers for bone scaffolds optimization. Polymers. 2020;12(4):905.

43. Park J, Lee SJ, Jung TG, Lee JH, Kim WD, Lee JY, et al. Surface modification of a three-dimensional polycaprolactone scaffold by polydopamine, biomineralization, and BMP-2 immobilization for potential bone tissue applications. Colloids and Surfaces B: Biointerfaces. 2021;199:111528.

44. Chen L, Shao L, Wang F, Huang Y, Gao F. Enhancement in sustained release of antimicrobial peptide and BMP-2 from degradable three dimensionalprinted PLGA scaffold for bone regeneration. Rsc Advances. 2019;9(19):10494–507.

45. Tateiwa D, Nakagawa S, Tsukazaki H, Okada R, Kodama J, Kushioka J, et al. A novel BMP-2–loaded



hydroxyapatite/beta-tricalcium phosphate microsphere/hydrogel composite for bone regeneration. Scientific reports. 2021;11(1):16924.

46. Chang H-C, Yang C, Feng F, Lin F-H, Wang C-H, Chang P-C. Bone morphogenetic protein-2 loaded poly (D, L-lactide-co-glycolide) microspheres enhance osteogenic potential of gelatin/hydroxyapatite/ β -tricalcium phosphate cryogel composite for alveolar ridge augmentation. Journal of the Formosan Medical Association. 2017;116(12):973–81.

47. Wang W, Zhou X, Wang H, Zhou G, Yu X. Fabrication and Evaluation of PCL/PLGA/ β -TCP Spiral-Structured Scaffolds for Bone Tissue Engineering. Bioengineering. 2024;11(7):732.

48. Mahmoud AH, Han Y, Dal-Fabbro R, Daghrery A, Xu J, Kaigler D, et al. Nanoscale β -TCP-laden GelMA/PCL composite membrane for guided bone regeneration. ACS applied materials & interfaces. 2023;15(27):32121–35.

49. Muzzio N, Moya S, Romero G. Multifunctional scaffolds and synergistic strategies in tissue engineering and regenerative medicine. Pharmaceutics. 2021;13(6):792.

50. He Y, Wang Q, Liu Y, Zhang Z, Cao Z, Wang S, et al. Composite mineralized collagen/polycaprolactone scaffold-loaded microsphere system with dual osteogenesis and antibacterial functions. Polymers. 2024;16(17):2394.

51. Kim Y, Lim JY, Yang GH, Seo J-H, Ryu H-S, Kim G. 3D-printed PCL/bioglass (BGS-7) composite scaffolds with high toughness and cell-responses for bone tissue regeneration. Journal of Industrial and Engineering Chemistry. 2019;79:163–71.

52. Gan Q, Zhu J, Yuan Y, Liu H, Qian J, Li Y, et al. A dual-delivery system of pH-responsive chitosan-functionalized mesoporous silica nanoparticles bearing BMP-2 and dexamethasone for enhanced bone regeneration. Journal of Materials Chemistry B. 2015;3(10):2056–66.

53. Gagnon D, Mouallem M, Leduc S, Rouleau DM, Chapleau J. A systematic scoping review of the latest data on orthobiologics in the surgical treatment of nonunion. Orthopaedics & Traumatology: Surgery & Research. 2024:103896.

54. Shah I, Chopra M, Ton A, Hah RJ, Wang JC, Alluri RK. Osteobiologics in degenerative spine disease: current clinical applications of recombinant human bone morphogenetic proteins—a narrative review. AME Medical Journal. 2024;9.

55. Govoni M, Vivarelli L, Mazzotta A, Stagni C, Maso A, Dallari D. Commercial bone grafts claimed as an alternative to autografts: Current trends for clinical applications in orthopaedics. Materials. 2021;14(12):3290.

56. Patel D, Tatum SA. Bone Graft Substitutes and Enhancement in Craniomaxillofacial Surgery. Facial Plastic Surgery. 2023;39(05):556–63.

57. Wang B, Feng C, Liu Y, Mi F, Dong J. Recent advances in biofunctional guided bone regeneration materials for repairing defective alveolar and maxillofacial bone: A review. Japanese Dental Science Review. 2022;58:233–48.

58. Xue P-P, Yuan J-d, Yao Q, Zhao Y-Z, Xu H-L. Bioactive factors-imprinted scaffold vehicles for promoting bone healing: The potential strategies and the confronted challenges for clinical production. BIO Integration. 2020;1(1):37–54.

59. Liu Q, Peng X, Liu X, Mou X, Guo Y, Yang L, et al. Advances in the application of bone morphogenetic proteins and their derived peptides in bone defect repair. Composites Part B: Engineering. 2023;262:110805.

60. Von Benecke JP, Tarsitano E, Zimmermann L-MA, Shakesheff KM, Walsh WR, Bae HW. A Narrative Review on Recombinant Human Bone Morphogenetic Protein 2: Where Are We Now? Cureus. 2024;16(8).

61. Chen X, Tan B, Bao Z, Wang S, Tang R, Wang Z, et al. Enhanced bone regeneration via spatiotemporal and controlled delivery of a genetically engineered BMP-2 in a composite Hydrogel. Biomaterials. 2021;277:121117.

62. Hasan LA. Biochemical Role of Bone Morphogenetic Protein and Oxidative Damage in the Rheumatoid Arthritis: College of Medicine Department of Chemistry and Biochemistry Biochemical ...; 2021.

63. Miller A. Complications of Bone Morphogenetic Protein-2 Use in Open Transforaminal Lumbar Interbody Fusion. 2021.

64. Afkhami E, Movaheddin M, Anvari M, Farashahi Yazd E, Aflatoonian B. Exosomes, a promising future for endometriosis. Regenerative Biomedicine. 2024;1(1):32– 5.

65. Karimi M, Izadi M, Akyash F, Aflatoonian B. Mesenchymal Stem/Stromal Cells Derived Exosomesbased Therapy and Polycystic Ovary Syndrome. Regenerative Biomedicine. 2025;1(2):135–8.

66. Karimi M, Anvari M, Mohammadi M, Hajizadeh-Tafti F, Aflatoonian B. Exosomes Extracted from Human Embryonic Stem Cells-Derived Mesenchymal Stem/Stromal Cells Support Treatment of Polycystic Ovary Syndrome in Animal Model. Regenerative Biomedicine. 2025;1(2):127–34.

67. Shokrollah N, Aflatoonian B, Ahmadieh-Yazdi A. Therapeutic Potential of Exosomes in Wound Healing with A Focus on Fibroblast-Derived Exosomes: A Mini Review. Regenerative Biomedicine. 2024;1(1):23–31.

68. Wang L, Jiang S, Zhou J, Gholipourmalekabadi M, Cao Y, Lin K, et al. From hard tissues to beyond: Progress and challenges of strontiumcontaining biomaterials in regenerative medicine applications. Bioactive Materials. 2025;49:85–120.

69. De Pace R, Molinari S, Mazzoni E, Perale G. Bone Regeneration: A Review of Current Treatment Strategies. Journal of Clinical Medicine. 2025;14(6):1838.

70. Gillman CE, Jayasuriya AC. FDA-approved bone grafts and bone graft substitute devices in bone regeneration. Materials Science and Engineering: C. 2021;130:112466.

71. Montemurro N, Pierozzi E, Inchingolo A, Pahwa B, De Carlo A, Palermo A, et al. New biograft solution, growth factors and bone regenerative approaches in neurosurgery, dentistry, and orthopedics: a review. European Review for Medical & Pharmacological Sciences. 2023;27(16).

72. Malham GM, Louie PK, Brazenor GA, Mobbs RJ, Walsh WR, Sethi RK. Recombinant human bone morphogenetic protein-2 in spine surgery: recommendations for use and alternative bone substitutes—a narrative review. Journal of Spine Surgery. 2022;8(4):477.

73. Liu S, Wang Y, Liang Z, Zhou M, Chen C. Comparative clinical effectiveness and safety of bone morphogenetic protein versus autologous iliac crest bone graft in lumbar fusion: a meta-analysis and systematic review. Spine. 2020;45(12):E729–E41.

74. On S-W, Park S-Y, Yi S-M, Park I-Y, Byun S-H, Yang B-E. Current status of recombinant human bone morphogenetic protein-2 (rhBMP-2) in maxillofacial surgery: should it be continued? Bioengineering. 2023;10(9):1005.

75. Papanna M, Al-Hadithy N, Somanchi B, Sewell M, Robinson P, Khan S, et al. The use of bone morphogenic protein-7 (OP-1) in the management of resistant non-unions in the upper and lower limb. Injury. 2012;43(7):1135–40.

76. Moghaddam A, Thaler B, Bruckner T, Tanner M, Schmidmaier G. Treatment of atrophic femoral nonunions according to the diamond concept: results of oneand two-step surgical procedure. Journal of orthopaedics. 2017;14(1):123–33.

77. Li G, Wu J, Cheng X, Pei X, Wang J, Xie W. Nanoparticle-Mediated Gene Delivery for Bone Tissue Engineering. Small. 2025;21(4):2408350.

78. Zhu X, Ma D, Yang B, An Q, Zhao J, Gao X, et al. Research progress of engineered mesenchymal stem cells and their derived exosomes and their application in autoimmune/inflammatory diseases. Stem cell research & therapy. 2023;14(1):71.

79. Tang G, Liu Z, Liu Y, Yu J, Wang X, Tan Z, et al. Recent trends in the development of bone regenerative biomaterials. Frontiers in Cell and Developmental biology. 2021;9:665813.

80. Bakhshandeh B, Ranjbar N, Abbasi A, Amiri E, Abedi A, Mehrabi MR, et al. Recent progress in the manipulation of biochemical and biophysical cues for engineering functional tissues. Bioengineering & Translational Medicine. 2023;8(2):e10383.

81. Tateiwa D, Kaito T. Advances in bone regeneration with growth factors for spinal fusion: A literature review. North American Spine Society Journal (NASSJ). 2023;13:100193.

82. Qin X, Xi Y, Jiang Q, Chen C, Yang G. Type H vessels in osteogenesis, homeostasis, and related disorders. Differentiation. 2023;134:20–30.

83. Packel L. Oncological diseases and disorders. Acute Care Physical Therapy. 2024;539–88.

84. Hye T, Hossain MR, Saha D, Foyez T, Ahsan F. Emerging biologics for the treatment of pulmonary arterial hypertension. Journal of drug targeting. 2023;31(5):471–85.

85. Shan BH, Wu FG. Hydrogel-based growth factor delivery platforms: strategies and recent advances. Advanced Materials. 2024;36(5):2210707.

86. Gu Z, Wang J, Fu Y, Pan H, He H, Gan Q, et al. Smart biomaterials for articular cartilage repair and regeneration. Advanced Functional Materials. 2023;33(10):2212561.

87. Bharathi R, Ganesh SS, Harini G, Vatsala K, Anushikaa R, Aravind S, et al. Chitosan-based scaffolds as drug delivery systems in bone tissue engineering. International Journal of Biological Macromolecules. 2022;222:132–53.

88. Tu Y, Li Y, Qu G, Ning Y, Li B, Li G, et al. A Review of Basic Fibroblast Growth Factor Delivery Strategies and Applications in Regenerative Medicine. Journal of Biomedical Materials Research Part A. 2025;113(1):e37834.

89. Wen C, Xu X, Zhang Y, Xia J, Liang Y, Xu L. Bone targeting nanoparticles for the treatment of osteoporosis. International Journal of Nanomedicine. 2024:1363–83.

90. Gao X, Ruzbarsky JJ, Layne JE, Xiao X, Huard J. Stem cells and bone tissue engineering. Life. 2024;14(3):287.

91. Han X, Alu A, Liu H, Shi Y, Wei X, Cai L, et al. Biomaterial-assisted biotherapy: A brief review of biomaterials used in drug delivery, vaccine development, gene therapy, and stem cell therapy. Bioactive materials. 2022;17:29–48.

92. Freitas GP, Lopes HB, Souza AT, Gomes MPO, Quiles GK, Gordon J, et al. Mesenchymal stem cells overexpressing BMP-9 by CRISPR-Cas9 present high in vitro osteogenic potential and enhance in vivo bone formation. Gene therapy. 2021;28(12):748–59.

93. Sivasankar M, Chinta ML, Rao PS. Zirconia based composite scaffolds and their application in bone tissue engineering. International Journal of Biological Macromolecules. 2024:130558.

94. Mo X, Zhang D, Liu K, Zhao X, Li X, Wang W. Nano-hydroxyapatite composite scaffolds loaded with bioactive factors and drugs for bone tissue engineering. International journal of molecular sciences. 2023;24(2):1291.

95. Sahebalzamani M, Ziminska M, McCarthy HO, Levingstone TJ, Dunne NJ, Hamilton AR. Advancing bone tissue engineering one layer at a time: a layer-bylayer assembly approach to 3D bone scaffold materials. Biomaterials Science. 2022;10(11):2734–58.

96. Shah SA, Sohail M, Nakielski P, Rinoldi C, Zargarian SS, Kosik-Kozioł A, et al. Integrating microand nanostructured platforms and biological drugs to enhance biomaterial-based bone regeneration strategies. Biomacromolecules. 2024;26(1):140–62.

97. Mahapatra C, Kumar P, Paul MK, Kumar A. Angiogenic stimulation strategies in bone tissue regeneration. Tissue and Cell. 2022;79:101908.

98. Tsiklin IL, Shabunin AV, Kolsanov AV, Volova LT. In vivo bone tissue engineering strategies: advances and prospects. Polymers. 2022 Aug 8;14(15):3222.

99. Kostadinova M, Raykovska M, Simeonov R, Lolov S, Mourdjeva M. Recent Advances in Bone Tissue Engineering: Enhancing the Potential of Mesenchymal Stem Cells for Regenerative Therapies. Current Issues in Molecular Biology. 2025 Apr 17;47(4):287.

100. Shu DY, Lovicu FJ. Insights into bone morphogenetic protein—(BMP-) signaling in ocular lens biology and pathology. Cells. 2021 Sep 30;10(10):2604.
101. Wang L, Ruan M, Bu Q, Zhao C. Signaling Pathways Driving MSC Osteogenesis: Mechanisms, Regulation, and Translational Applications. International Journal of Molecular Sciences. 2025 Feb 4;26(3):1311.

102. Jang HJ, Yoon JK. The role of vasculature and angiogenic strategies in bone regeneration. Biomimetics. 2024 Jan 26;9(2):75.

2024 Jan 26;9(2):75. 103. Novais A, Chatzopoulou E, Chaussain C, Gorin C. The potential of FGF-2 in craniofacial bone tissue engineering: A review. Cells. 2021 Apr 17;10(4):932.

104. Galarraga-Vinueza ME, Barootchi S, Nevins ML, Nevins M, Miron RJ, Tavelli L. Twenty-five years of recombinant human growth factors rhPDGF-BB and rhBMP-2 in oral hard and soft tissue regeneration. Periodontology 2000. 2024 Feb;94(1):483-509.

105. Zhu L, Liu Y, Wang A, Zhu Z, Li Y, Zhu C, Che Z, Liu T, Liu H, Huang L. Application of BMP in bone tissue engineering. Frontiers in Bioengin

eering and Biotechnology. 2022 Mar 31;10:810880. 106. Percival KM, Paul V, Husseini GA. Recent advancements in bone tissue engineering: integrating smart scaffold technologies and bio-responsive systems for enhanced regeneration. International Journal of Molecular Sciences. 2024 May 30;25(11):6012.

107. Jiang S, Wang M, He J. A review of biomimetic scaffolds for bone regeneration: toward a cell-free strategy. Bioengineering & Translational Medicine. 2021 May;6(2):e10206.

108. Zhou W, Yan K, Xi Q. BMP signaling in cancer stemness and differentiation. Cell Regeneration. 2023 Dec 5;12(1):37.

109. Wu M, Chen G, Li YP. TGF- β and BMP signaling in osteoblast, skeletal development, and bone formation, homeostasis and disease. Bone research. 2016 Apr 26;4(1):1-21.

110. Reddy LV, Murugan D, Mullick M, Begum Moghal ET, Sen D. Recent approaches for angiogenesis in search of successful tissue engineering and regeneration. Current stem cell research & therapy. 2020 Feb 1;15(2):111-34.

111. Rose L, Fitzsimmons R, El-Bialy T, Uludağ H. Gene Therapy in Bone Regeneration: A Summary of Delivery Approaches for Effective Therapies. Intracellular Delivery: Fundamentals and Applications. 2011:813-46.

